Estrogens and Anti-Estrogens: Key Mediators of Prostate Carcinogenesis and New Therapeutic Candidates

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Abstract Despite the historical use of estrogens in the treatment of prostate cancer (PCa) little is known about their direct biological effects on the prostate, their role in carcinogenesis, and what mechanisms mediate their therapeutic effects on PCa. It is now known that estrogens alone, or in synergism with an androgen, are potent inducers of aberrant growth and neoplastic transformation in the prostate. The mechanisms of estrogen carcinogenicity could be mediated via induction of unscheduled cell proliferation or through metabolic activation of estrogens to genotoxic metabolites. Agerelated changes and race-/ethnic-based differences in circulating or locally formed estrogens may explain differential PCa risk among different populations. Loss of expression of estrogen receptor (ER)- β expression during prostate carcinogenesis and prevention of estrogen-mediated oxidative damage could be exploited in future PCa prevention strategies. Reexpression of ER-β in metastatic PCa cells raises the possibility of using ER-β-specific ligands in triggering cell death in these malignant cells. A variety of new estrogenic/anti-estrogenic/selective estrogen receptor modulator (SERM)-like compounds, including 2-methoxyestradiol, genistein, resveratrol, licochalcone, Raloxifene, ICI 182,780, and estramustine are being evaluated for their potential in the next generation of PCa therapies. Increasing numbers of patients selfmedicate with herbal formulations such as PC-SPES. Some of these compounds are selective ER- β ligands, while most of them have minimal interaction with ER-a. Although many may inhibit testosterone production by blockade of the hypothalamal-pituitary-testis axis, the most effective agents also exhibit direct cytostatic, cytotoxic, or apoptotic action on PCa cells. Some of them are potent in interfering with tubulin polymerization, blocking angiogenesis and cell motility, suppressing DNA synthesis, and inhibiting specific kinase activities. Further discovery of other compounds with potent apoptotic activities but minimal estrogen action should promote development of a new generation of effective PCa preventive or treatment regimens with few or no side-effects due to estrogenicity. Further advancement of our knowledge of the role of estrogens in prostate carcinogenesis through metabolic activation of estrogens and/or ER-mediated pathways will certainly result in better preventive or therapeutic modalities for PCa. J. Cell. Biochem. 91: 491–503, 2004. © 2003 Wiley-Liss, Inc.

Key words: estrogen receptor; anti-estrogens; selective estrogen receptor modulator (SERM); estrogen receptor relates; estrogen receptor co-regulators; oxidative stress; genomic damages; prostate cancer risk; hormone refractory disease; hormonal therapy

Historically, androgens have been considered to be the major sex hormones that regulate normal and malignant growth of the prostate [Ho et al., 1997; Bosland, 2000; Prins, 2000; Taplin and Ho, 2001; Risbridger et al., 2003]. However, recent evidence suggests that estrogens significantly contribute to normal prostatic functions as well as to the genesis of

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prostate cancer (PCa). Paradoxically, diethylstilbestrol (DES), a xenoestrogen, was used as the first standard therapy for PCa treatment [Huggins and Hodges, 1941]. Recently, novel phytoestrogens [Castle and Thrasher, 2002; Morrissey and Watson, 2003] and synthetic estrogens/anti-estrogens/selective estrogen receptor modulators (SERMs) [Steiner et al., 2001; Kim et al., 2002; Steiner and Raghow, 2003] have emerged as promising PCa preventive and treatment agents. The primary objective of this review is to address the mechanisms of action of estrogen carcinogenicity in the prostate in order to enrich our understanding of the roles played by these hormones in the pathogenesis of PCa. The review is also

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intended to evaluate the potential of a new generation of estrogenic and anti-estrogenic compounds as therapies for the prevention and treatment of PCa.

Estrogens/Anti-Estrogens as Contributing Factors to the Genesis and Progression of PCa

It is well established that estrogens play important roles in normal growth, differentiation, and development of the prostate [Ho et al., 1997; Bosland, 2000; Prins, 2000; Taplin and Ho, 2001; Risbridger et al., 2003]. However, any significant contribution by these steroids to the development of prostatic diseases, including PCa in men and animals, is not yet apparent. Estrogens alone, or in synergism with androgens, are potent inducers of aberrant growth and neoplastic transformation in the prostate. Administration of pharmacological doses of estrogen induces a well-known proliferative lesion, termed squamous metaplasia, in the prostates of various mammalian species, including humans [Leav et al., 1978; Merk et al., 1982, 1986; Goodwin and Cummings, 1984; Sugimura et al., 1988; Levine et al., 1991; Yonemura et al., 1995; Cunha et al., 2001; Risbridger et al., 2001]. The metaplastic change is initiated by the proliferation of basal epithelial cells, which subsequently differentiate into squamous cells. In Noble rats, administration of estrogens in conjunction with an androgen leads to the development of a pre-cancerous lesion similar to human prostatic intraepithelial neoplasia (PIN) and a high incidence of full blown prostate adenocarcinoma [Noble, 1977, 1980; Drago et al., 1979; Leav et al., 1988, 1989; Ofner et al., 1992; Bosland et al., 1995; Ho and Lane, 1996; Wong et al., 1998; Tam et al., 2000]. The combined hormonal treatment also initiated and promoted malignant transformation in Rb-deficient prostatic tissues grafted under the kidney capsules of immune-deficient mice [Wang et al., 2000].

Epidemiological findings reveal that African– Americans, who have the highest incidence of PCa in the USA, also have significantly higher levels of serum estrogens, even at a young age, when compared with their Caucasian– American peers [Hill et al., 1984; Ross et al., 1986; Henderson et al., 1988; de Jong et al., 1991; Bosland, 2000]. In contrast, lower circulating estrogen levels have been noted in Japanese men, who are known to have a low risk of PCa when compared with a higher-risk group, Caucasian–Dutch men [de Jong et al., 1991]. Furthermore, the incidence of PCa rises exponentially in elderly men, in whom the ratio of estrogen to androgen may increase by up to 40%. This age-related hormonal shift is due primarily to a decline in testicular function and an increase in aromatization of adrenal androgens by peripheral adipose tissue during aging [Vermeulen et al., 1972; Zumoff et al., 1982; Gray et al., 1991a,b; Griffiths, 2000]. At the cellular level, significant declines in 5α -dihydrotestosterone (DHT) levels and increases in estrogen (E2 and estrone) levels have been observed in prostatic epithelial cells [Krieg et al., 1993]. Collectively, these endocrine changes at mid-life, termed andropause, may significantly augment estrogenic stimulation of the prostatic epithelium leading to reactivation of growth and subsequent neoplastic transformation. Several theories have been advanced to explain the mechanism of action of estrogen carcinogenicity in the prostate. The first suggests that enhancement of unscheduled cell proliferation under rising estrogenic influence may lead to the accumulation of spontaneous genetic errors in progenitor cells [Henderson and Feigelson, 2000]. Others propose direct and/or indirect estrogen- or estrogen metabolite-mediated genotoxicity as causative factors of neoplastic transformation of the prostatic epithelium.

Estrogen Metabolism as a Determinant of Prostatic Carcinogenesis

In female tissues like the breast and endometrium, estrogens induce tumors partially via metabolic activation of the natural estrogens to genotoxic metabolites, such as catechol estrogens and quinone/semiguinone intermediates [Cavalieri et al., 2000; Yager, 2000]. These metabolites induce genomic damage directly by formation of DNA adducts or indirectly via formation of reactive oxygen species (ROS), which can cause genomic and protein damages and lipid peroxidation. Conversion of estrogens to 2-hydroxy (OH)-estrogens is primarily mediated by the enzyme cytochrome P450 1A1 (gene: CYP1A1) and into 4-OH-estrogens by cytochrome P450 1B1 (gene: CYP1B1) [Yager and Liehr, 1996]. As an important cellular defensive mechanism, these highly reactive catechol estrogens are inactivated quickly by the enzyme catechol-O-methyl-transferase (COMT). Additionally, cells are equipped with several isoforms of glutathione-S-transferases (GSTs) that detoxify ROS and other electrophilic moieties by conjugative reactions. Genetic polymorphisms that alter enzyme activity and DNA hypermethylation- or mutation-mediated silencing of key estrogen metabolizing genes have been associated with higher or lower risks for breast and endometrial cancer [Yager, 2000]. In other words, the levels and activities of these enzymes collectively influence the magnitude and the duration of lifetime exposure to these potentially transforming estrogen metabolites. Although many of the aforementioned pathways are well characterized in female estrogen-sensitive organs, few have been investigated for their contribution to prostate carcinogenesis. Dorsolateral prostate carcinomas arise in 100% of Noble rats treated with estradiol and testosterone. It was demonstrated that the combined hormone treatment induced specific DNA adduct formation in prostatic epithelial cells of the proximal ducts, where malignant lesions are believed to originate [Han et al., 1995]. Increased DNA single-strand breaks [Ho and Roy, 1994], elevated lipid peroxidation [Ho and Roy, 1994], accumulation of 8-oxo-2(-deoxyguanosine and nitrated proteins [Tam et al., unpublished data], down regulation of antioxidant defense enzyme activities [Tam et al., 2003], and accumulation of 4-OH-estradiol and its quinone derivatives [Cavalieri et al., 2002] were also noted in the cancer-susceptible dorsolateral prostates of animals treated with estrogen plus testosterone. Importantly, evidence of reduced activities of COMT and quinone reductase observed in the dorsolateral prostates of the treated rats might explain the preferential accumulation of these transforming estrogen metabolites in the rat prostate.

Recent molecular epidemiological data indicate that Japanese men homozygous for the G allele of SNP2455A > G of CYP1A1, a polymorphism that confers higher enzyme activity, were more susceptible to PCa development [Murata et al., 2001]. However, in an American population that was 86% Caucasian, this polymorphism was associated with a lower PCa risk [Chang et al., 2003]. The discrepancy could be due to ethnic/racial differences or other factors yet to be identified. In the same population, three other polymorphisms in the CYP1A1 gene were found to be associated with increased PCa risk [Chang et al., 2003]. Considering the important roles played by CYP1A1 in the oxidative metabolism of estrogen, the impact of CYP1A1 polymorphisms on CaP risk is worthy of further investigation. Of equal importance is a recent finding of higher PCa risk in Japanese men carrying a polymorphism at codon 119 (homozygous T/T compared to G/G 453) of CYP1B1 [Tanaka et al., 2002]. This polymorphism is believed to confer longer halflife to the enzyme CYP1B1, which converts estrogens into 4-OH estrogens. It has been postulated that 4-OH-estrogens are more carcinogenic than 2-OH-estrogens in female organs such as the breast [Yager, 2000]. A similar mechanism has yet to be identified in the prostate. Another gap in our understanding of the connection between estrogen metabolism and the pathogenesis of PCa is our lack of data on the enzyme COMT, which can convert 2-OH-catecholestrogens to 2-methoxyestradiol. Since 2methoxyestradiol has recently been shown to exhibit potent apoptotic activity against rapidly growing PCa cells [Lakhani et al., 2003], it would be of importance to correlate expression levels and polymorphisms of *COMT* to PCa risk in future epidemiological studies. Similarly, in situ production of estrogen from androgen via the activity of the enzyme aromatase has not been fully investigated as a risk factor modulator of PCa. Both aromatase protein and its enzyme activity have been demonstrated in PCa and benign prostatic hyperplasia (BPH) [Matzkin and Soloway, 1992; Di Salle et al., 1994; Hiramatsu et al., 1997]. Unfortunately, most clinical trials testing aromatase inhibitors have focused on their efficacy in treating BPH [Radlmaier et al., 1996; Suzuki et al., 1998] and attention has not been given to their use in PCa prevention or treatment. Studies of transgenic mice overexpressing aromatase (AROM+) [Li et al., 2001] and aromatase knockout mice (ArKO) [McPherson et al., 2001] have yielded only limited insights into the relationship between in situ estrogen formation and PCa risk. This is largely due to the fact that numerous endocrine factors are altered in these animals, which makes data interpretation difficult. Since it has been shown that alternative use of promoter to transactivate aromatase is an important determinant of breast carcinogenesis [Harada et al., 1993; Chen et al., 1999], studies focusing on establishing a similar scenario should be conducted for PCa. Finally, a recent observational study has reported that obesity, commonly associated with more aromatization of androgens to estrogen, is associated with a poorer histology in young men with PCa, especially men younger than 50 years of age [Rohrmann et al., 2003]. This finding is in concordance with the hypothesis that in situ production of estrogen via aromatization may contribute to tumor initiation and progression in the prostate.

Estrogen Receptors (ERs) as Functionally Divergent Mediators of Estrogen Action

Until recently the effects of estrogens on all tissues, including the prostate, were believed to be mediated by a steroid hormone receptor, now termed ER- α [Nilsson and Gustafsson, 2000; Pettersson and Gustafsson, 2001]. In 1996, another ER subtype, referred to as ER- β , was identified in rats [Kuiper et al., 1996] and in humans [Mosselman et al., 1996]. The two ER subtypes are structurally similar, consisting of the six common domains (A-F) found in all steroid hormone receptors. The DNA-binding domain (DBD) of ER- β , e.g., differs from ER- α by only three amino acids. In contrast, the Nterminal A/B domain of both receptors share only a 15.5% homology, the ligand binding domains (LBDs) are 58% homologous, and the F domains are distinctly different [Kuiper et al., 1996: Mosselman et al., 1996: Nilsson and Gustafsson, 2000; Pettersson and Gustafsson, 2001]. These structural dissimilarities are thought to reflect functional differences between the two receptors. ER- α and ER- β have been shown to bind to the same ligand with different affinities [Kuiper et al., 1997]. For example, genistein and some SERMs are known to bind ER- β with higher affinities than to ER- α . In addition, after binding to estrogens/antiestrogens/SERMs, the two ERs may use different enhancer elements such as the estrogen responsive element (ERE), the activating protein-1 (AP-1) site, the cyclic AMP-response element, the antioxidant responsive element, and the NF-kappa B site in the promoter regions of various genes [Paech et al., 1997; Cerillo et al., 1998; Montano et al., 1998; Pelzer et al., 2001; Weatherman et al., 2001; Liu et al., 2002]. It is now believed that the two ER-subtypes could mediate diverse biological functions through interaction with specific ligands and activation of different enhancer elements/transcriptional factor binding sites. Adding to the complexity of mechanisms mediating ER action is the

involvement of co-regulators that are required for ER signaling. The possibility exists that the two ER subtypes, depending on their binding ligand, interact with different coactivators, corepressors, and integrators to achieve the broadest degree of functional diversity and specificity [Osborne et al., 2001; Tremblay and Giguere, 2002].

Expression of ER Subtypes in Normal and Malignant Prostates: Implications in Tumor Initiation and Progression

The precise biological functions of the two ER subtypes in the prostate are currently undefined and their possible role in PCa initiation and progression is likewise unknown. Several studies have described the expression of the receptors, at both mRNA and protein levels, in the epithelial and stromal compartments of the normal and malignant adult human prostate [Horvath et al., 2001; Latil et al., 2001; Leav et al., 2001; Royuela et al., 2001; Torlakovic et al., 2002; Weihua et al., 2002; Fixemer et al., 2003; Tsurusaki et al., 2003]. ER- β is localized predominantly to the basal epithelial cell compartment of the normal human prostate, where ER- α is rarely found. ER- α is mainly expressed in the stroma of the normal gland. Results from these studies suggest that ER- β may exert a protective effect against aberrant cell proliferation and/or carcinogenesis [Chang and Prins, 1999; Poelzl et al., 2000; Signoretti and Loda, 2001]. Furthermore, findings in ER-β knockout mice indicate that these animals develop prostatic hyperplasia in old age, a phenomenon that does not occur in ER- α knockout mice [Krege et al., 1998]. A protective role of ER- β against proliferative lesions is also suggested by the studies of Prins [Chang and Prins, 1999]. Neonatal exposure of rats to estrogen causes downregulation of ER- β , upregulation of ER- α , and development of hyperplastic, dysplastic, and neoplastic lesions in the adult ventral prostates. Other suggested functions of ER- β in the normal prostate are related to its role in protecting tissues against oxidative injury. It has been shown that the ER- β is more effective than ER- α in transactivation of the electrophile/ antioxidant response element and therefore is a better inducer of antioxidant enzymes such as quinone reductase and GST [Montano and Katzenellenbogen, 1997; Montano et al., 2000]. This action of the ER- β , though not yet demonstrated in the prostate, is known to mediate artheroprotective [Dimitrova et al., 2002] and neuroprotective [Treuter et al., 2000; Mize et al., 2003] effects of estrogens.

In a comprehensive study [Leav et al., 2001] the expression of and rogen receptor (AR), ER- α , and ER- β in dyplastic lesions, primary carcinomas and in metastases of PCa was compared with that in normal glands from archival specimens. In the peripheral zone of the normal prostate, ER- β was overwhelmingly localized in the nuclei of basal cells, the purported proliferative compartment of the human prostate [De Marzo et al., 1998]. In the stroma of the peripheral zone, both ER- α and ER- β were expressed, while AR was present in both stromal and differentiated secretory cells. Compared with the normal epithelium, downregulation of ER- β was observed in high-grade PIN lesions, while the expression of ER- α or AR in these lesions remained unchanged. Interestingly, ER- β was strongly expressed in grade 3 carcinoma but was markedly diminished in carcinomas that were Gleason grade 4/5. Additional studies [Horvath et al., 2001; Latil et al., 2001; Pasquali et al., 2001a,b; Tsurusaki et al., 2003] have confirmed a loss or reduced expression of ER- β as PCa progresses in the primary site. These findings suggest a role for ER- β in limiting basal cells proliferation [McNeal, 1988: Bonkhoff et al., 1994: McNeal et al., 1995], which may help preserve genomic integrity [Signoretti and Loda, 2001; Weihua et al., 2002]. In addition, the proposed involvement of ER- β in regulating antioxidant enzyme expression [Montano and Katzenellenbogen, 1997; Montano et al., 2000] may protect basal cells from oxidative damage, which is a risk factor for neoplastic transformation. In this regard, the strong expression of ER- β in basal epithelial cells may be necessary to counteract androgen's stimulation of continuous growth [Lee, 1997] and/or its pro-oxidant action [Ripple et al., 1997, 1999]. Disruption of this equilibrium may trigger aberrant growth and transformation in the progenitor cells within the prostate basal epithelial layer, which has been postulated to harbor susceptible cell populations that give rise to PCa [Garraway et al., 2003].

In marked contrast, ER- β protein but not ER- α protein was dramatically expressed in the majority of metastases of PCa in bone and regional lymph nodes [Leav et al., 2001]. It is currently unclear why ER- β expression is elevated in metastases. Either local factors in the

distant metastatic sites may induce ER- β expression or a clonal selection process may occur in primary tumors. The latter scenario would argue that PCa cells that fail to loss ER- β expression (for unknown reasons) have higher matastatic potentials and, therefore, escape to distant sites. Adding to the complexity of understanding the ER- β regulation of PCa development, a recent study [Fujimura et al., 2001] has found that while wild-type ER- β expression was significantly lower in the cancers than in the benign epithelium, ER-βcx (a C-terminal truncated splice variant of ER- β) was expressed at significantly higher levels in high-grade cancers (83%) compared with the low-grade tumors (22%) and the benign tissues. Although the functional roles of ER-Bcx remain unclear, these findings raise the possibility that wild-type ER- β and ER- β cx are independent predictors of PCa. All in all, it has become apparent that ER- β is a key determinant of the genesis, progression and metastasis of PCa. Additional research is needed to fully unveil its contribution to the initiation and progression of PCa.

Finally, ER- α , expressed mainly in the stromal compartment of the prostate, may also contribute to the pathogenesis of PCa. In a study of the genotypic and allelic frequencies of the six different polymorphic loci of *ER*- α in a Japanese population, polymorphism in codon 10 of *ER*- α was found to be a possible risk factor for PCa [Tanaka et al., 2003].

Estrogen Imprinting and PCa Risks

Experimental and epidemiological evidence suggest that PCa risk in adulthood could be determined by exposure to estrogens during embryonic, perinatal/neonatal, or peripubertal development, a phenomenon referred to as "estrogen imprinting" [Rajfer and Coffey, 1978]. Perinatal and neonatal exposure of rats [Arai et al., 1977, 1983; Vorherr et al., 1979; Prins, 1992] or mice [McLachlan et al., 1975; McLachlan, 1977; Pylkkanen et al., 1991] to estrogens or estrogen mimics induces development of various proliferative lesions in the adult gland. These changes may be partially explained by permanent changes in androgen receptor levels, ER statuses, interacinar stromal tissue volume, and proliferative potential of the prostatic epithelium [Prins, 1992; vom Saal et al., 1997; Prins et al., 1998; Prins et al., 2001]. The "estrogenized" adult glands also demonstrate a lack of differentiation, disorganization of the acini, and excessive luminal sloughing. Analogous to these animal studies, it has recently been reported that the fetal human prostate does not express ER- α at the protein level but does strongly express ER-^β [Adams et al., 2002]. This finding indicates that ER- β is the sole transducer of estrogen action in the fetal human prostate during in utero development. Observation studies provide additional support for a fetal base determination of adult PCa risk. It has been shown that circulating E2 levels of pregnant African-American women are 37% higher than those of European-American women [Henderson et al., 1988]. Furthermore, Ekbom and associates [Ekbom et al., 1996, 2000] have reported that indicators of high levels of endogenous pregnancy hormones, such as high birth weight and jaundice in the offspring, are associated with increased risk for PCa, while indicators of low levels, like pre-eclampsia, are associated with decreased risk [Ekbom et al., 1996]. If intrauterine or perinatal environments are important for PCa risk in adulthood, cancer prevention strategies should be directed to these early developmental stages.

SERMs, Phytoestrogens, and new Estrogenic/Anti-Estrogenic Agents as Potential Therapies for PCa

The hypothesis that estrogen inhibits growth of PCa originates in the classic studies of Huggins and Hodges [1941] who pioneered the use of a potent xenoestrogen, DES, to treat men with PCa. Castration combined with DES was proven to be an affordable, highly efficient treatment, but it quickly fell out of favor due to cardiac toxicity [Cox and Crawford, 1995; Denis and Griffiths, 2000; Taplin and Ho, 2001]. A few clinical trials have found that the first generation of anti-estrogens/SERMs, such as tamoxifen and toremifene, as alternatives to DES, are not effective therapies [Bergan et al., 1999; Stein et al., 2001]. Despite these early set-backs, the likelihood of developing more effective and less toxic SERMs/estrogen-like compounds to treat PCa treatment has improved substantially, coincident with our growing knowledge on the differential expression of ER subtypes and differential action of estrogen/anti-estrogen/SERM in normal and malignant prostatic epithelial cells.

Traditionally, the primary mechanism by which estrogens are believed to induce PCa regression is through action on the hypothalamic-pituitary axis, thereby inhibiting testosterone synthesis [El Rayes and Hussain, 2002]. However, it is now known that many estrogens/ anti-estrogens/phytoestrogens/SERMs including DES, 2-methoxy-E₂, genistein, resveratrol, licochalcone, Raloxifene, ICI 182,780, and estramustine have antitumor effects independent of this pathway. Their ability to suppress PCa cell growth has been attributed to a broad range of actions including direct cytotoxicity [Schulz et al., 1988], interruption of cell cycle progression [Kumar et al., 2001; Qadan et al., 2001], induction of apoptosis [Qadan et al., 2001; Kim et al., 2002; Shimada et al., 2003], depolymerization of microtubules [Dahllof et al., 1993], inhibition of DNA synthesis [Kuwajerwala et al., 2002], inhibition of topoisomerase II [Matsukawa et al., 1993], blockade of tyrosine kinase [Matsukawa et al., 1993; Misra et al., 2002], disruption of apoptotic regulators such as bcl-2 [Rafi et al., 2000], and activation of death domain receptors [Mor et al., 2000; LaVallee et al., 2003]. The ER subtype responsible for mediating the anti-tumor action of the pure anti-estrogen ICI-182,780 [Lau et al., 2000], the SERM Raloxifene [Kim et al., 2002], the phytoestrogen genistein, and others [An et al., 2001; Boue et al., 2003] is likely ER- β and not ER- α .

In addition to their ability to induce cell cycle arrest/apoptosis in PCa cells, some of these estrogenic/anti-estrogenic compounds are also potent inhibitors of angiogenesis and metastasis [Li and Sarkar, 2002]. Genistein was shown to upregulate expression of genes with antiangiogenesis or anti-metastasis properties [Li and Sarkar, 2002] and to activate the cell adhesion-associated signaling via focal adhesion kinase [Liu et al., 2000] in PCa cells. Raloxifene, a SERM, administered to tumorbearing male Lobund–Wistar (LW) rats, was shown to produce marked reduction in PCa metastasis to lymphatic and pulmonary tissues [Neubauer et al., 1995].

Estramustine, when combined with a variety of microtubular inhibitors, has clinically been proven highly active in treating hormonerefractory PCa [Gilligan and Kantoff, 2002]. Several new estrogenic/anti-estrogenic compounds are now in clinical phase I and phase II studies for PCa prevention or treatment. These include 2-methoxyestradiol [Lakhani et al., 2003], genistein [Miltyk et al., 2003], clover-derived dietary isoflavones [Jarred et al., 2002], and two new SERMs, GTx-006 and arzoxifene [Steiner et al., 2001]. The hope is that these novel estrogens/anti-estrogens, phytoestrogens or SERMs can selectively activate apoptosis or cell cycle arrest in rapidly growing PCa cells, via non-ER- α and/or -ER- β signaling pathways, offering highly effective therapies for PCa while avoiding some of the undesirable side-effects believed to be caused by the "classical" estrogen action.

Herbal Estrogenic Formulations in Prevention and Treatment of PCA

Complementary and alternative medicines (CAMs) are estimated to be used by 7-64% of PCa patients, with younger and more educated men more likely to use them [Kao and Devine, 2000; Porterfield, 2000; Steginga et al., 2001]. Among CAM therapies for PCa, PC-SPES is by far the most widely used due to its potency [Kao and Devine, 2000; Yip et al., 2003]. It has recently been recalled by the US Food and Drug Administration because of contamination with the synthetic estrogen DES [Guns et al., 2002]. PC-SPES is an herbal formulation previously available as a dietary supplement sold in 'natural food' or 'health food' stores in the US. It is comprised of extracts from eight different plants: chrysanthemum. Isatis indigotica, Glycyrrhiza glabra, Ganoderma lucidum, Panax pseudo-ginseng, Rabdosia rebescens, saw palmetto, and scutellaria baicalensis [Thomson et al., 2002]. A handful of clinical studies indicate that the formulation has good clinical efficacy and benefits most patients (97%) who have hormonally-responsive PCa [Small et al., 2000; Pirani, 2001; Oh and Small, 2002]. It also has activity against a fraction of hormoneindependent PCa [de la et al., 2000; Pfeifer et al., 2000; Small et al., 2000; Oh et al., 2001]. Experimental studies have documented in vitro and in vivo efficacy of the formulation and described possible estrogenic mechanisms of action [Hsieh et al., 1997, 1998; DiPaola et al., 1998; Kubota et al., 2000]. Biochemical studies indicated that PC-SPES has potent estrogenic activity similar to 1 nM estradiol. However, mass spectrometry analysis indicated that the estrogenic effect was not due to DES, estrone, or E2 [Hsieh et al., 1997, 1998; Darzynkiewicz et al., 2000]. The unsupervised use of herbal CAM by so many has raised serious concerns. First, the specific active estrogenic agents

responsible for the therapeutic efficacy have not been isolated or characterized. Secondly, there is a paucity of data to demonstrate convincingly the safety, efficacy, effectiveness, and mechanisms of the effects of these compounds. Most importantly, up to 72% of PCa patients do not inform their physicians [Lippert et al., 1999] when they are using these therapies, while they are also receiving standard treatments for PCa. This may adversely affect or confound statistics surrounding the outcomes of standard treatments [Kao and Devine, 2000]. Finally, the difference in philosophies between Western medicine and herbal medicine may make evaluation of herbal medicine difficult when the same standards established to test synthetic pharmaceutical compounds were to be used as criteria. In order to advance CAM usage, welldesigned clinical trials, perhaps using a set of modified standards, are necessary to assess the efficacy and safety of CAMs in PCa treatment [Yip et al., 2003]. Unfortunately, such trials are expected to be highly complex and expensive, and will require lengthy follow-up. Until such trials are completed, our knowledge and understanding of the mechanisms of action and efficacies of CAMs in PCa prevention and treatment will continue to be sketchy [Wilkinson and Chodak, 2003].

Overall Conclusions

The central role of estrogens in PCa development has been well supported by epidemiological findings and experimental animal data. Although it has been demonstrated in female estrogen-sensitive organs that estrogens can act as genotoxic carcinogens through metabolic activation, the importance of estrogen metabolites in prostate carnogenesis has thus far received very little attention. Data from animal studies support the hypothesis that formation of genotoxic metabolites and reactive intermediates as a probable cause of PCa initiation. Individuals expressing specific polymorphisms of estrogen metabolizing enzymes and local production of estrogen via aromatization probably determine the lifetime exposure and influence PCa risks among different ethnic/ racial/geographical groups. The recent discoverv of ER- β , in addition to ER- α , provides new insights to the mechanism(s) of ERmediated events associated with PCa initiation and progression. Availability of a host of new estrogenic/anti-estrogenic/SERM-like compounds and unearthing of novel anticancer actions afforded by these compounds on PCa cells open avenues for the development of a new generation of PCa therapies with high antitumor potency and minimal systemic sideeffects caused by their innate estrogenicity.

REFERENCES

- Adams JY, Leav I, Lau KM, Ho SM, Pflueger SM. 2002. Expression of estrogen receptor beta in the fetal, neonatal, and prepubertal human prostate. Prostate 52: 69–81.
- An J, Tzagarakis-Foster C, Scharschmidt TC, Lomri N, Leitman DC. 2001. Estrogen receptor beta-selective transcriptional activity and recruitment of coregulators by phytoestrogens. J Biol Chem 276:17808-17814.
- Arai Y, Suzuki Y, Nishizuka Y. 1977. Hyperplastic and metaplastic lesions in the reproductive tract of male rats induced by neonatal treatment with diethylstilbestrol. Virchows Arch A Pathol Anat Histol 376:21–28.
- Arai Y, Mori T, Suzuki Y, Bern HA. 1983. Long-term effects of perinatal exposure to sex steroids and diethylstilbestrol on the reproductive system of male mammals. Int Rev Cytol 84:235–268.
- Bergan RC, Reed E, Myers CE, Headlee D, Brawley O, Cho HK, Figg WD, Tompkins A, Linehan WM, Kohler D, Steinberg SM, Blagosklonny MV. 1999. A Phase II study of high-dose tamoxifen in patients with hormone-refractory prostate cancer. Clin Cancer Res 5:2366–2373.
- Bonkhoff H, Stein U, Remberger K. 1994. The proliferative function of basal cells in the normal and hyperplastic human prostate. Prostate 24:114–118.
- Bosland MC. 2000. The role of steroid hormones in prostate carcinogenesis. J Natl Cancer Inst Monogr 27:39–66.
- Bosland MC, Ford H, Horton L. 1995. Induction at high incidence of ductal prostate adenocarcinomas in NBL/Cr and Sprague–Dawley Hsd:SD rats treated with a combination of testosterone and estradiol-17 beta or diethylstilbestrol. Carcinogenesis 16:1311–1317.
- Boue SM, Wiese TE, Nehls S, Burow ME, Elliott S, Carter-Wientjes CH, Shih BY, McLachlan JA, Cleveland TE. 2003. Evaluation of the estrogenic effects of legume extracts containing phytoestrogens. J Agric Food Chem 51:2193-2199.
- Castle EP, Thrasher JB. 2002. The role of soy phytoestrogens in prostate cancer. Urol Clin North Am 29:71-ix.
- Cavalieri E, Frenkel K, Liehr JG, Rogan E, Roy D. 2000. Estrogens as endogenous genotoxic agents—DNA adducts and mutations. J Natl Cancer Inst Monogr 27: 75–93.
- Cavalieri EL, Devanesan P, Bosland MC, Badawi AF, Rogan EG. 2002. Catechol estrogen metabolites and conjugates in different regions of the prostate of Noble rats treated with 4-hydroxyestradiol: Implications for estrogen-induced initiation of prostate cancer. Carcinogenesis 23:329–333.
- Cerillo G, Rees A, Manchanda N, Reilly C, Brogan I, White A, Needham M. 1998. The oestrogen receptor regulates NFkappaB and AP-1 activity in a cell-specific manner. J Steroid Biochem Mol Biol 67:79–88.

- Chang WY, Prins GS. 1999. Estrogen receptor-beta: Implications for the prostate gland. Prostate 40:115– 124.
- Chang BL, Zheng SL, Isaacs SD, Turner A, Hawkins GA, Wiley KE, Bleecker ER, Walsh PC, Meyers DA, Isaacs WB, Xu J. 2003. Polymorphisms in the *CYP1A1* gene are associated with prostate cancer risk. Int J Cancer 106: 375–378.
- Chen S, Zhou D, Okubo T, Kao YC, Yang C. 1999. Breast tumor aromatase: Functional role and transcriptional regulation. Endocr Relat Cancer 6:149–156.
- Cox RL, Crawford ED. 1995. Estrogens in the treatment of prostate cancer. J Urol 154:1991–1998.
- Cunha GR, Wang YZ, Hayward SW, Risbridger GP. 2001. Estrogenic effects on prostatic differentiation and carcinogenesis. Reprod Fertil Dev 13:285–296.
- Dahllof B, Billstrom A, Cabral F, Hartley-Asp B. 1993. Estramustine depolymerizes microtubules by binding to tubulin. Cancer Res 53:4573-4581.
- Darzynkiewicz Z, Traganos F, Wu JM, Chen S. 2000. Chinese herbal mixture PC SPES in treatment of prostate cancer (review). Int J Oncol 17:729-736.
- de Jong FH, Oishi K, Hayes RB, Bogdanowicz JF, Raatgever JW, van der Maas PJ, Yoshida O, Schroeder FH. 1991. Peripheral hormone levels in controls and patients with prostatic cancer or benign prostatic hyperplasia: Results from the Dutch–Japanese case-control study. Cancer Res 51:3445–3450.
- de la TA, Buttyan R, Hayek O, Bagiella E, Shabsigh A, Burchardt M, Burchardt T, Chopin DK, Katz AE. 2000. Herbal therapy PC-SPES: In vitro effects and evaluation of its efficacy in 69 patients with prostate cancer. J Urol 164:1229-1234.
- De Marzo AM, Meeker AK, Epstein JI, Coffey DS. 1998. Prostate stem cell compartments: Expression of the cell cycle inhibitor p27Kip1 in normal, hyperplastic, and neoplastic cells. Am J Pathol 153:911–919.
- Denis LJ, Griffiths K. 2000. Endocrine treatment in prostate cancer. Semin Surg Oncol 18:52-74.
- Di Salle E, Briatico G, Giudici D, Ornati G, Panzeri A. 1994. Endocrine properties of the testosterone 5 alpha-reductase inhibitor turosteride (FCE 26073). J Steroid Biochem Mol Biol 48:241–248.
- Dimitrova KR, DeGroot KW, Suyderhoud JP, Pirovic EA, Munro TJ, Wieneke J, Myers AK, Kim YD. 2002. 17-beta estradiol preserves endothelial cell viability in an in vitro model of homocysteine-induced oxidative stress. J Cardiovasc Pharmacol 39:347–353.
- DiPaola RS, Zhang H, Lambert GH, Meeker R, Licitra E, Rafi MM, Zhu BT, Spaulding H, Goodin S, Toledano MB, Hait WN, Gallo MA. 1998. Clinical and biologic activity of an estrogenic herbal combination (PC-SPES) in prostate cancer. N Engl J Med 339:785–791.
- Drago JR, Ikeda RM, Maurer RE, Goldman LB, Tesluk H. 1979. The Nb rat: Prostatic adenocarcinoma model. Invest Urol 16:353–359.
- Ekbom A, Hsieh CC, Lipworth L, Wolk A, Ponten J, Adami HO, Trichopoulos D. 1996. Perinatal characteristics in relation to incidence of and mortality from prostate cancer. BMJ 313:337–341.
- Ekbom A, Wuu J, Adami HO, Lu CM, Lagiou P, Trichopoulos D, Hsieh C. 2000. Duration of gestation and prostate cancer risk in offspring. Cancer Epidemiol Biomarkers Prev 9:221–223.

- El Rayes BF, Hussain MH. 2002. Hormonal therapy for prostate cancer: Past, present and future. Expert Rev Anticancer Ther 2:37–47.
- Fixemer T, Remberger K, Bonkhoff H. 2003. Differential expression of the estrogen receptor beta (ERbeta) in human prostate tissue, premalignant changes, and in primary, metastatic, and recurrent prostatic adenocarcinoma. Prostate 54:79–87.
- Fujimura T, Takahashi S, Urano T, Ogawa S, Ouchi Y, Kitamura T, Muramatsu M, Inoue S. 2001. Differential expression of estrogen receptor beta (ERbeta) and its Cterminal truncated splice variant ERbetacx as prognostic predictors in human prostatic cancer. Biochem Biophys Res Commun 289:692–699.
- Garraway LA, Lin D, Signoretti S, Waltregny D, Dilks J, Bhattacharya N, Loda M. 2003. Intermediate basal cells of the prostate: In vitro and in vivo characterization. Prostate 55:206–218.
- Gilligan T, Kantoff PW. 2002. Chemotherapy for prostate cancer. Urology 60:94–100.
- Goodwin WE, Cummings RH. 1984. Squamous metaplasia of the verumontanum with obstruction due to hypertrophy: Long-term effects of estrogen on the prostate in an aging male-to-female transsexual. J Urol 131: 553–554.
- Gray A, Berlin JA, McKinlay JB, Longcope C. 1991a. An examination of research design effects on the association of testosterone and male aging: Results of a metaanalysis. J Clin Epidemiol 44:671–684.
- Gray A, Feldman HA, McKinlay JB, Longcope C. 1991b. Age, disease, and changing sex hormone levels in middleaged men: Results of the Massachusetts Male Aging Study. J Clin Endocrinol Metab 73:1016–1025.
- Griffiths K. 2000. Estrogens and prostatic disease. International Prostate Health Council Study Group. Prostate 45:87–100.
- Guns ES, Goldenberg SL, Brown PN. 2002. Mass spectral analysis of PC-SPES confirms the presence of diethylstilbestrol. Can J Urol 9:1684–1688.
- Han X, Liehr JG, Bosland MC. 1995. Induction of a DNA adduct detectable by ³²P-postlabeling in the dorsolateral prostate of NBL/Cr rats treated with estradiol-17 beta and testosterone. Carcinogenesis 16: 951-954.
- Harada N, Utsumi T, Takagi Y. 1993. Tissue-specific expression of the human aromatase cytochrome P-450 gene by alternative use of multiple exons 1 and promoters, and switching of tissue-specific exons 1 in carcinogenesis. Proc Natl Acad Sci USA 90:11312-11316.
- Henderson BE, Feigelson HS. 2000. Hormonal carcinogenesis. Carcinogenesis 21:427–433.
- Henderson BE, Bernstein L, Ross RK, Depue RH, Judd HL. 1988. The early in utero oestrogen and testosterone environment of Blacks and Whites: Potential effects on male offspring. Br J Cancer 57:216–218.
- Hill P, Garbaczewski L, Walker AR. 1984. Age, environmental factors and prostatic cancer. Med Hypotheses 14:29–39.
- Hiramatsu M, Maehara I, Ozaki M, Harada N, Orikasa S, Sasano H. 1997. Aromatase in hyperplasia and carcinoma of the human prostate. Prostate 31:118–124.
- Ho SM, Lane K. 1996. Sex hormone-induction and dietary modulation of prostatic adenocarcinoma (PA) in animal models. Urol Onco 110–115.

- Ho SM, Roy D. 1994. Sex hormone-induced nuclear DNA damage and lipid peroxidation in the dorsolateral prostates of Noble rats. Cancer Lett 84:155–162.
- Ho SM, Lane K, Lee K. 1997. Neoplastic transformation of the prostate. In: Naz RK, editor. Prostate: Basic and clinical aspects. New York CRC Press. pp 74–114.
- Horvath LG, Henshall SM, Lee CS, Head DR, Quinn DI, Makela S, Delprado W, Golovsky D, Brenner PC, O'Neill G, Kooner R, Stricker PD, Grygiel JJ, Gustafsson JA, Sutherland RL. 2001. Frequent loss of estrogen receptorbeta expression in prostate cancer. Cancer Res 61:5331– 5335.
- Hsieh T, Chen SS, Wang X, Wu JM. 1997. Regulation of androgen receptor (AR) and prostate specific antigen (PSA) expression in the androgen-responsive human prostate LNCaP cells by ethanolic extracts of the Chinese herbal preparation, PC-SPES. Biochem Mol Biol Int 42:535–544.
- Hsieh TC, Ng C, Chang CC, Chen SS, Mittleman A, Wu JM. 1998. Induction of apoptosis and down-regulation of bcl-6 in mutu I cells treated with ethanolic extracts of the Chinese herbal supplement PC-SPES. Int J Oncol 13: 1199–1202.
- Huggins C, Hodges C. 1941. Studies on prostatic cancer. I. The effect of castration, of estrogen and androgen injection on serum phosphatases in metastatic carcinoma of the prostate. Cancer Res 1:293–298.
- Jarred RA, Keikha M, Dowling C, McPherson SJ, Clare AM, Husband AJ, Pedersen JS, Frydenberg M, Risbridger GP. 2002. Induction of apoptosis in low to moderategrade human prostate carcinoma by red clover-derived dietary isoflavones. Cancer Epidemiol Biomarkers Prev 11:1689–1696.
- Kao GD, Devine P. 2000. Use of complementary health practices by prostate carcinoma patients undergoing radiation therapy. Cancer 88:615–619.
- Kim IY, Kim BC, Seong DH, Lee DK, Seo JM, Hong YJ, Kim HT, Morton RA, Kim SJ. 2002. Raloxifene, a mixed estrogen agonist/antagonist, induces apoptosis in androgen-independent human prostate cancer cell lines. Cancer Res 62:5365–5369.
- Krege JH, Hodgin JB, Couse JF, Enmark E, Warner M, Mahler JF, Sar M, Korach KS, Gustafsson JA, Smithies O. 1998. Generation and reproductive phenotypes of mice lacking estrogen receptor beta. Proc Natl Acad Sci USA 95:15677–15682.
- Krieg M, Nass R, Tunn S. 1993. Effect of aging on endogenous level of 5 alpha-dihydrotestosterone, testosterone, estradiol, and estrone in epithelium and stroma of normal and hyperplastic human prostate. J Clin Endocrinol Metab 77:375–381.
- Kubota T, Hisatake J, Hisatake Y, Said JW, Chen SS, Holden S, Taguchi H, Koeffler HP. 2000. PC-SPES: A unique inhibitor of proliferation of prostate cancer cells in vitro and in vivo. Prostate 42:163–171.
- Kuiper GG, Enmark E, Pelto-Huikko M, Nilsson S, Gustafsson JA. 1996. Cloning of a novel receptor expressed in rat prostate and ovary. Proc Natl Acad Sci USA 93:5925–5930.
- Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, Gustafsson JA. 1997. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. Endocrinology 138:863–870.

- Kumar AP, Garcia GE, Slaga TJ. 2001. 2-Methoxyestradiol blocks cell-cycle progression at G(2)/M phase and inhibits growth of human prostate cancer cells. Mol Carcinog 31:111–124.
- Kuwajerwala N, Cifuentes E, Gautam S, Menon M, Barrack ER, Reddy GP. 2002. Resveratrol induces prostate cancer cell entry into s phase and inhibits DNA synthesis. Cancer Res 62:2488–2492.
- Lakhani NJ, Sarkar MA, Venitz J, Figg WD. 2003. 2-Methoxyestradiol, a promising anticancer agent. Pharmacotherapy 23:165–172.
- Latil A, Bieche I, Vidaud D, Lidereau R, Berthon P, Cussenot O, Vidaud M. 2001. Evaluation of androgen, estrogen (ER alpha and ER beta), and progesterone receptor expression in human prostate cancer by realtime quantitative reverse transcription-polymerase chain reaction assays. Cancer Res 61:1919-1926.
- Lau KM, LaSpina M, Long J, Ho SM. 2000. Expression of estrogen receptor (ER)-alpha and ER-beta in normal and malignant prostatic epithelial cells: Regulation by methylation and involvement in growth regulation. Cancer Res 60:3175–3182.
- LaVallee TM, Zhan XH, Johnson MS, Herbstritt CJ, Swartz G, Williams MS, Hembrough WA, Green SJ, Pribluda VS. 2003. 2-Methoxyestradiol up-regulates death receptor 5 and induces apoptosis through activation of the extrinsic pathway. Cancer Res 63:468–475.
- Leav I, Merk FB, Ofner P, Goodrich G, Kwan PW, Stein BM, Sar M, Stumpf WE. 1978. Bipotentiality of response to sex hormones by the prostate of castrated or hypophysectomized dogs. Direct effects of estrogen. Am J Pathol 93:69-92.
- Leav I, Ho SM, Ofner P, Merk FB, Kwan PW, Damassa D. 1988. Biochemical alterations in sex hormone-induced hyperplasia and dysplasia of the dorsolateral prostates of Noble rats. J Natl Cancer Inst 80:1045–1053.
- Leav I, Merk FB, Kwan PW, Ho SM. 1989. Androgensupported estrogen-enhanced epithelial proliferation in the prostates of intact Noble rats. Prostate 15:23–40.
- Leav I, Lau KM, Adams JY, McNeal JE, Taplin ME, Wang J, Singh H, Ho SM. 2001. Comparative studies of the estrogen receptors beta and alpha and the androgen receptor in normal human prostate glands, dysplasia, and in primary and metastatic carcinoma. Am J Pathol 159:79–92.
- Lee C. 1997. Cellular interactions in prostate cancer. Br J Urol 79(Suppl 1):21–27.
- Levine AC, Kirschenbaum A, Droller M, Gabrilove JL. 1991. Effect of the addition of estrogen to medical castration on prostatic size, symptoms, histology and serum prostate specific antigen in 4 men with benign prostatic hypertrophy. J Urol 146:790-793.
- Li Y, Sarkar FH. 2002. Gene expression profiles of genistein-treated PC3 prostate cancer cells. J Nutr 132:3623-3631.
- Li X, Nokkala E, Yan W, Streng T, Saarinen N, Warri A, Huhtaniemi I, Santti R, Makela S, Poutanen M. 2001. Altered structure and function of reproductive organs in transgenic male mice overexpressing human aromatase. Endocrinology 142:2435–2442.
- Lippert MC, McClain R, Boyd JC, Theodorescu D. 1999. Alternative medicine use in patients with localized prostate carcinoma treated with curative intent. Cancer 86:2642-2648.

- Liu Y, Kyle E, Lieberman R, Crowell J, Kellof G, Bergan RC. 2000. Focal adhesion kinase (FAK) phosphorylation is not required for genistein-induced FAK-beta-1-integrin complex formation. Clin Exp Metastasis 18: 203–212.
- Liu MM, Albanese C, Anderson CM, Hilty K, Webb P, Uht RM, Price RH, Jr., Pestell RG, Kushner PJ. 2002. Opposing action of estrogen receptors alpha and beta on cyclin D1 gene expression. J Biol Chem 277:24353– 24360.
- Matsukawa Y, Marui N, Sakai T, Satomi Y, Yoshida M, Matsumoto K, Nishino H, Aoike A. 1993. Genistein arrests cell cycle progression at G2-M. Cancer Res 53:1328-1331.
- Matzkin H, Soloway MS. 1992. Response to second-line hormonal manipulation monitored by serum PSA in stage D2 prostate carcinoma. Urology 40:78–80.
- McLachlan JA. 1977. Prenatal exposure to diethylstilbestrol in mice: Toxicological studies. J Toxicol Environ Health 2:527–537.
- McLachlan JA, Newbold RR, Bullock B. 1975. Reproductive tract lesions in male mice exposed prenatally to diethylstilbestrol. Science 190:991–992.
- McNeal JE. 1988. Normal histology of the prostate. Am J Surg Pathol 12:619–633.
- McNeal JE, Haillot O, Yemoto C. 1995. Cell proliferation in dysplasia of the prostate: Analysis by PCNA immunostaining. Prostate 27:258–268.
- McPherson SJ, Wang H, Jones ME, Pedersen J, Iismaa TP, Wreford N, Simpson ER, Risbridger GP. 2001. Elevated androgens and prolactin in aromatase-deficient mice cause enlargement, but not malignancy, of the prostate gland. Endocrinology 142:2458–2467.
- Merk FB, Ofner P, Kwan PW, Leav I, Vena RL. 1982. Ultrastructural and biochemical expressions of divergent differentiation in prostates of castrated dogs treated with estrogen and androgen. Lab Invest 47:437–450.
- Merk FB, Warhol MJ, Kwan PW, Leav I, Alroy J, Ofner P, Pinkus GS. 1986. Multiple phenotypes of prostatic glandular cells in castrated dogs after individual or combined treatment with androgen and estrogen. Morphometric, ultrastructural, and cytochemical distinctions. Lab Invest 54:442–456.
- Miltyk W, Craciunescu CN, Fischer L, Jeffcoat RA, Koch MA, Lopaczynski W, Mahoney C, Jeffcoat RA, Crowell J, Paglieri J, Zeisel SH. 2003. Lack of significant genotoxicity of purified soy isoflavones (genistein, daidzein, and glycitein) in 20 patients with prostate cancer. Am J Clin Nutr 77:875–882.
- Misra RR, Hursting SD, Perkins SN, Sathyamoorthy N, Mirsalis JC, Riccio ES, Crowell JA. 2002. Genotoxicity and carcinogenicity studies of soy isoflavones. Int J Toxicol 21:277–285.
- Mize AL, Shapiro RA, Dorsa DM. 2003. Estrogen receptormediated neuroprotection from oxidative stress requires activation of the mitogen-activated protein kinase pathway. Endocrinology 144:306–312.
- Montano MM, Katzenellenbogen BS. 1997. The quinone reductase gene: A unique estrogen receptor-regulated gene that is activated by antiestrogens. Proc Natl Acad Sci USA 94:2581–2586.
- Montano MM, Jaiswal AK, Katzenellenbogen BS. 1998. Transcriptional regulation of the human quinone reductase gene by antiestrogen-liganded estrogen

receptor-alpha and estrogen receptor-beta. J Biol Chem 273:25443–25449.

- Montano MM, Wittmann BM, Bianco NR. 2000. Identification and characterization of a novel factor that regulates quinone reductase gene transcriptional activity. J Biol Chem 275:34306–34313.
- Mor G, Kohen F, Garcia-Velasco J, Nilsen J, Brown W, Song J, Naftolin F. 2000. Regulation of fas ligand expression in breast cancer cells by estrogen: Functional differences between estradiol and tamoxifen. J Steroid Biochem Mol Biol 73:185–194.
- Morrissey C, Watson RW. 2003. Phytoestrogens and prostate cancer. Curr Drug Targets 4:231–241.
- Mosselman S, Polman J, Dijkema R. 1996. ER beta: Identification and characterization of a novel human estrogen receptor. FEBS Lett 392:49–53.
- Murata M, Watanabe M, Yamanaka M, Kubota Y, Ito H, Nagao M, Katoh T, Kamataki T, Kawamura J, Yatani R, Shiraishi T. 2001. Genetic polymorphisms in cytochrome P450 (CYP) 1A1, CYP1A2, CYP2E1, glutathione Stransferase (GST) M1 and GSTT1 and susceptibility to prostate cancer in the Japanese population. Cancer Lett 165:171–177.
- Neubauer BL, Best KL, Counts DF, Goode RL, Hoover DM, Jones CD, Sarosdy MF, Shaar CJ, Tanzer LR, Merriman RL. 1995. Raloxifene (LY156758) produces antimetastatic responses and extends survival in the PAIII rat prostatic adenocarcinoma model. Prostate 27:220–229.
- Nilsson S, Gustafsson JA. 2000. Estrogen receptor transcription and transactivation: Basic aspects of estrogen action. Breast Cancer Res 2:360–366.
- Noble RL. 1977. The development of prostatic adenocarcinoma in Nb rats following prolonged sex hormone administration. Cancer Res 37:1929–1933.
- Noble RL. 1980. Production of Nb rat carcinoma of the dorsal prostate and response of estrogen-dependent transplants to sex hormones and tamoxifen. Cancer Res 40:3547-3550.
- Ofner P, Bosland MC, Vena RL. 1992. Differential effects of diethylstilbestrol and estradiol-17 beta in combination with testosterone on rat prostate lobes. Toxicol Appl Pharmacol 112:300–309.
- Oh WK, Small EJ. 2002. Complementary and alternative therapies in prostate cancer. Semin Oncol 29:575–584.
- Oh WK, George DJ, Hackmann K, Manola J, Kantoff PW. 2001. Activity of the herbal combination, PC-SPES, in the treatment of patients with androgen-independent prostate cancer. Urology 57:122–126.
- Osborne CK, Schiff R, Fuqua SA, Shou J. 2001. Estrogen receptor: Current understanding of its activation and modulation. Clin Cancer Res 7:4338s-4342s.
- Paech K, Webb P, Kuiper GG, Nilsson S, Gustafsson J, Kushner PJ, Scanlan TS. 1997. Differential ligand activation of estrogen receptors ERalpha and ERbeta at AP1 sites. Science 277:1508–1510.
- Pasquali D, Rossi V, Esposito D, Abbondanza C, Puca GA, Bellastella A, Sinisi AA. 2001a. Loss of estrogen receptor beta expression in malignant human prostate cells in primary cultures and in prostate cancer tissues. J Clin Endocrinol Metab 86:2051–2055.
- Pasquali D, Staibano S, Prezioso D, Franco R, Esposito D, Notaro A, De Rosa G, Bellastella A, Sinisi AA. 2001b. Estrogen receptor beta expression in human prostate tissue. Mol Cell Endocrinol 178:47–50.

- Pelzer T, Neumann M, de Jager T, Jazbutyte V, Neyses L. 2001. Estrogen effects in the myocardium: Inhibition of NF-kappaB DNA binding by estrogen receptor-alpha and -beta. Biochem Biophys Res Commun 286:1153–1157.
- Pettersson K, Gustafsson JA. 2001. Role of estrogen receptor beta in estrogen action. Annu Rev Physiol 63:165–192.
- Pfeifer BL, Pirani JF, Hamann SR, Klippel KF. 2000. PC-SPES, a dietary supplement for the treatment of hormone-refractory prostate cancer. BJU Int 85: 481-485.
- Pirani JF. 2001. The effects of phytotherapeutic agents on prostate cancer: An overview of recent clinical trials of PC SPES. Urology 58:36–38.
- Poelzl G, Kasai Y, Mochizuki N, Shaul PW, Brown M, Mendelsohn ME. 2000. Specific association of estrogen receptor beta with the cell cycle spindle assembly checkpoint protein, MAD2. Proc Natl Acad Sci USA 97:2836-2839.
- Porterfield H. 2000. UsToo PC-SPES surveys: Review of studies and update of previous survey results. Mol Urol 4:289–291.
- Prins GS. 1992. Neonatal estrogen exposure induces lobespecific alterations in adult rat prostate androgen receptor expression. Endocrinology 130:3703–3714.
- Prins GS. 2000. Molecular biology of the androgen receptor. Mayo Clin Proc 75(Suppl):S32–S35.
- Prins GS, Sklarew RJ, Pertschuk LP. 1998. Image analysis of androgen receptor immunostaining in prostate cancer accurately predicts response to hormonal therapy. J Urol 159:641–649.
- Prins GS, Birch L, Couse JF, Choi I, Katzenellenbogen B, Korach KS. 2001. Estrogen imprinting of the developing prostate gland is mediated through stromal estrogen receptor alpha: Studies with alphaERKO and betaERKO mice. Cancer Res 61:6089–6097.
- Pylkkanen L, Santti R, Newbold R, McLachlan JA. 1991. Regional differences in the prostate of the neonatally estrogenized mouse. Prostate 18:117–129.
- Qadan LR, Perez-Stable CM, Anderson C, D'Ippolito G, Herron A, Howard GA, Roos BA. 2001. 2-Methoxyestradiol induces G2/M arrest and apoptosis in prostate cancer. Biochem Biophys Res Commun 285:1259–1266.
- Radlmaier A, Eickenberg HU, Fletcher MS, Fourcade RO, Reis Santos JM, van Aubel OG, Bono AV. 1996. Estrogen reduction by aromatase inhibition for benign prostatic hyperplasia: Results of a double-blind, placebo-controlled, randomized clinical trial using two doses of the aromatase-inhibitor atamestane. Atamestane Study Group. Prostate 29:199–208.
- Rafi MM, Rosen RT, Vassil A, Ho CT, Zhang H, Ghai G, Lambert G, DiPaola RS. 2000. Modulation of bcl-2 and cytotoxicity by licochalcone-A, a novel estrogenic flavonoid. Anticancer Res 20:2653–2658.
- Rajfer J, Coffey DS. 1978. Sex steroid imprinting of the immature prostate. Long-term effects. Invest Urol 16: 186-190.
- Ripple MO, Henry WF, Rago RP, Wilding G. 1997. Prooxidant-antioxidant shift induced by androgen treatment of human prostate carcinoma cells. J Natl Cancer Inst 89:40–48.
- Ripple MO, Hagopian K, Oberley TD, Schatten H, Weindruch R. 1999. Androgen-induced oxidative stress in human LNCaP prostate cancer cells is associated with

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multiple mitochondrial modifications. Antioxid Redox Signal 1:71-81.

- Risbridger GP, Wang H, Frydenberg M, Cunha G. 2001. The metaplastic effects of estrogen on mouse prostate epithelium: Proliferation of cells with basal cell phenotype. Endocrinology 142:2443-2450.
- Risbridger GP, Bianco JJ, Ellem SJ, McPherson SJ. 2003. Oestrogens and prostate cancer. Endocr Relat Cancer 10:187–191.
- Rohrmann S, Roberts WW, Walsh PC, Platz EA. 2003. Family history of prostate cancer and obesity in relation to high-grade disease and extraprostatic extension in young men with prostate cancer. Prostate 55: 140-146.
- Ross R, Bernstein L, Judd H, Hanisch R, Pike M, Henderson B. 1986. Serum testosterone levels in healthy young Black and White men. J Natl Cancer Inst 76: 45-48.
- Royuela M, de Miguel MP, Bethencourt FR, Sanchez-Chapado M, Fraile B, Arenas MI, Paniagua R. 2001. Estrogen receptors alpha and beta in the normal, hyperplastic and carcinomatous human prostate. J Endocrinol 168:447-454.
- Schulz P, Bauer HW, Brade WP, Keller A, Fittler F. 1988. Evaluation of the cytotoxic activity of diethylstilbestrol and its mono- and di-phosphate towards prostatic carcinoma cells. Cancer Res 48:2867–2870.
- Shimada K, Nakamura M, Ishida E, Kishi M, Konishi N. 2003. Requirement of c-jun for testosterone-induced sensitization to N-(4-hydroxyphenyl)retinamide-induced apoptosis. Mol Carcinog 36:115–122.
- Signoretti S, Loda M. 2001. Estrogen receptor beta in prostate cancer: Brake pedal or accelerator? Am J Pathol 159:13–16.
- Small EJ, Frohlich MW, Bok R, Shinohara K, Grossfeld G, Rozenblat Z, Kelly WK, Corry M, Reese DM. 2000. Prospective trial of the herbal supplement PC-SPES in patients with progressive prostate cancer. J Clin Oncol 18:3595–3603.
- Steginga SK, Occhipinti S, Dunn J, Gardiner RA, Heathcote P, Yaxley J. 2001. The supportive care needs of men with prostate cancer (2000). Psychooncology 10:66–75.
- Stein S, Zoltick B, Peacock T, Holroyde C, Haller D, Armstead B, Malkowicz SB, Vaughn DJ. 2001. Phase II trial of toremifene in androgen-independent prostate cancer: A Penn cancer clinical trials group trial. Am J Clin Oncol 24:283–285.
- Steiner MS, Raghow S. 2003. Antiestrogens and selective estrogen receptor modulators reduce prostate cancer risk. World J Urol 21:31–36.
- Steiner MS, Raghow S, Neubauer BL. 2001. Selective estrogen receptor modulators for the chemoprevention of prostate cancer. Urology 57:68–72.
- Sugimura Y, Cunha GR, Yonemura CU, Kawamura J. 1988. Temporal and spatial factors in diethylstilbestrolinduced squamous metaplasia of the developing human prostate. Hum Pathol 19:133–139.
- Suzuki K, Okazaki H, Ono Y, Kurokawa K, Suzuki T, Onuma E, Takanashi H, Mamiya Y, Yamanaka H. 1998. Effect of dual inhibition of 5-alpha-reductase and aromatase on spontaneously developed canine prostatic hypertrophy. Prostate 37:70–76.
- Tam NN, Chung SS, Lee DT, Wong YC. 2000. Aberrant expression of hepatocyte growth factor and its receptor,

c-Met, during sex hormone-induced prostatic carcinogenesis in the Noble rat. Carcinogenesis 21:2183– 2191.

- Tam NN, Ghatak S, Ho SM. 2003. Sex hormone-induced alterations in the activities of antioxidant enzymes and lipid peroxidation status in the prostate of Noble rats. Prostate 55:1–8.
- Tanaka Y, Sasaki M, Kaneuchi M, Shiina H, Igawa M, Dahiya R. 2002. Polymorphisms of the *CYP1B1* gene have higher risk for prostate cancer. Biochem Biophys Res Commun 296:820–826.
- Tanaka Y, Sasaki M, Kaneuchi M, Shiina H, Igawa M, Dahiya R. 2003. Polymorphisms of estrogen receptor alpha in prostate cancer. Mol Carcinog 37:202–208.
- Taplin ME, Ho SM. 2001. Clinical review 134: The endocrinology of prostate cancer. J Clin Endocrinol Metab 86:3467-3477.
- Thomson JO, Dzubak P, Hajduch M. 2002. Prostate cancer and the food supplement, PC-SPES, minireview. Neoplasma 49:69–74.
- Torlakovic E, Lilleby W, Torlakovic G, Fossa SD, Chibbar R. 2002. Prostate carcinoma expression of estrogen receptor-beta as detected by PPG5/10 antibody has positive association with primary Gleason grade and Gleason score. Hum Pathol 33:646-651.
- Tremblay GB, Giguere V. 2002. Coregulators of estrogen receptor action. Crit Rev Eukaryot Gene Expr 12: 1–22.
- Treuter E, Warner M, Gustafsson JA. 2000. Mechanism of oestrogen signalling with particular reference to the role of ER beta in the central nervous system. Novartis Found Symp 230:7–14.
- Tsurusaki T, Aoki D, Kanetake H, Inoue S, Muramatsu M, Hishikawa Y, Koji T. 2003. Zone-dependent expression of estrogen receptors alpha and beta in human benign prostatic hyperplasia. J Clin Endocrinol Metab 88:1333– 1340.
- Vermeulen A, Rubens R, Verdonck L. 1972. Testosterone secretion and metabolism in male senescence. J Clin Endocrinol Metab 34:730-735.
- vom Saal FS, Timms BG, Montano MM, Palanza P, Thayer KA, Nagel SC, Dhar MD, Ganjam VK, Parmigiani S, Welshons WV. 1997. Prostate enlargement in mice due to fetal exposure to low doses of estradiol or diethylstilbestrol and opposite effects at high doses. Proc Natl Acad Sci USA 94:2056–2061.
- Vorherr H, Messer RH, Vorherr UF, Jordan SW, Kornfeld M. 1979. Teratogenesis and carcinogenesis in rat offspring after transplacental and transmammary exposure to diethylstilbestrol. Biochem Pharmacol 28: 1865–1877.
- Wang Y, Hayward SW, Donjacour AA, Young P, Jacks T, Sage J, Dahiya R, Cardiff RD, Day ML, Cunha GR. 2000. Sex hormone-induced carcinogenesis in Rb-deficient prostate tissue. Cancer Res 60:6008–6017.
- Weatherman RV, Clegg NJ, Scanlan TS. 2001. Differential SERM activation of the estrogen receptors (ERalpha and ERbeta) at AP-1 sites. Chem Biol 8:427–436.
- Weihua Z, Warner M, Gustafsson JA. 2002. Estrogen receptor beta in the prostate. Mol Cell Endocrinol 193:1-5.
- Wilkinson S, Chodak GW. 2003. Critical review of complementary therapies for prostate cancer. J Clin Oncol 21:2199–2210.

- Wong YC, Wang YZ, Tam NN. 1998. The prostate gland and prostate carcinogenesis. Ital J Anat Embryol 103:237–252.
- Yager JD. 2000. Endogenous estrogens as carcinogens through metabolic activation. J Natl Cancer Inst Monogr 27:67–73.
- Yager JD, Liehr JG. 1996. Molecular mechanisms of estrogen carcinogenesis. Annu Rev Pharmacol Toxicol 36:203-232.
- Yip I, Cudiamat M, Chim D. 2003. PC-SPES for treatment of prostate cancer: Herbal medicine. Curr Urol Rep 4:253-257.
- Yonemura CY, Cunha GR, Sugimura Y, Mee SL. 1995. Temporal and spatial factors in diethylstilbestrol-induced squamous metaplasia in the developing human prostate. II. Persistent changes after removal of diethylstilbestrol. Acta Anat (Basel) 153: 1-11.
- Zumoff B, Levin J, Strain GW, Rosenfeld RS, O'Connor J, Freed SZ, Kream J, Whitmore WS, Fukushima DK, Hellman L. 1982. Abnormal levels of plasma hormones in men with prostate cancer: Evidence toward a "twodisease" theory. Prostate 3:579–588.